

## CRISPR/cas9 mediated gene knockout in mosquito vector *Aedes aegypti*: *In-silico* approach

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*Aedes aegypti* is the principal vector of several viruses including Dengue, Chikungunya and yellow fever. The recent advent of Clustered-Regularly-Interspaced-Short-Palindromic-Repeats (CRISPR)-Cas9 has sparked significant enthusiasm for the genetic control of *A. aegypti*. In the current study, we hypothesized that CRISPR-cas9 mediated gene knockout of doublesex (*Aedsex*) and Actin-4 (*AeAct-4*) in *A. aegypti* would result in sterile intersex phenotype and flightless phenotype respectively in females. Designing effective sgRNA is the most critical part of CRISPR/Cas9 editing. Therefore, sgRNAs were designed *in-silico* with the aim of developing efficient measures to control *A. aegypti* populations. Gene sequences of *Aedsex* (IAAEL009114) and *AeAct-4* (AAEL001951) were retrieved from sequence data available in the VectorBase database. BLASTP was performed in searching for paralogs. Phylogenetic analysis was performed for all paralogs with  $\geq 80\%$  amino acid similarity. Eight paralogs were identified for *the AeAct-4* while none was identified for *Aedsex*. DNA sequences for the paralogs of *AeAct-4* were aligned to identify targets for sgRNAs that were unique to *AeAct-4*. Exon 2 was selected for *AeAct-4*, as the target site. For the *Aedsex* gene, 5a and 5b female-specific exons were selected as the target sequence for *AedsexF1* and *AedsexF2* isoforms respectively. sgRNAs were designed using online tools CHOP CHOP and CRISPR guide express. Two sgRNAs for each *AeAct-4* gene, *AedsexF1* and *AedsexF2* isoforms with minimum off-target effects and *in-silico* predicted efficiency  $> 60\%$  were selected to maximise the on-target gene knockout *in-vivo*. The efficiency of gene knockout can be evaluated by microinjecting synthesized sgRNAs directly into mosquito embryos with Cas9 protein.

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